ayny

Criofolinine and Vernavosine, New Pentacyclic Indole Alkaloids Incorporating Pyrroloazepine and Pyridopyrimidine Moieties Derived from a Common Yohimbine Precursor

Choy-Eng Nge,† Chew-Yan Gan,‡ Kuan-Hon Lim,§ Kang-Nee Ting,[∥] Yun-Yee Low,† and Toh-Seok Kam*,†

† Department of Chemis[try,](#page-3-0) Faculty of Science, University of Malaya, 50603 Kuala Lumpur, Malaysia ‡ University Malaysia of Computer Science and Engineering, Jalan Alamanda 2, Precint 1, 62000 Putrajaya, Malaysia §
§School of Pharmacy and [∥]Department of Biomedical Sciences, University of Nottingham Malaysia Campus, Jalan Broga, 43500 Semenyih, Selangor, Malaysia

S Supporting Information

[AB](#page-3-0)STRACT: [Two new in](#page-3-0)dole alkaloids characterized by previously unencountered natural product skeletons, viz., criofolinine (1), incorporating a pyrroloazepine motif within a pentacyclic ring system, and vernavosine (2, isolated as its ethyl ether derivative 3, which on hydrolysis regenerated the putative precursor alkaloid 2), incorporating a pyridopyrimidine moiety embedded within a pentacyclic carbon framework, were isolated from a Malayan Tabernaemontana species. The

structures and absolute configuration of these alkaloids were determined on the basis of NMR and MS analysis and confirmed by X-ray diffraction analysis.

 \sum lants belonging to the genus *Tabernaemontana* have a pantropical distribution,¹ are rich in alkaloids,² and continue to provide alkaloids with diverse structures and a continue to provide alkaloids with diverse structures and a wide range of biological activ[it](#page-3-0)ies.<su[p](#page-3-0)>2−9</sup> We previously reported the isolation and structure determination of two monoterpenoid indole alkaloids, voatinggin[e](#page-3-0) [an](#page-3-0)d tabertinggine, characterized by previously unknown natural product carbon skeletons derived from a common cleavamine-type precursor, from a Malayan Tabernaemontana.⁹ We now report the isolation and structure determination of two additional minor alkaloids, representing first members [o](#page-3-0)f new structural groups of the monoterpenoid indoles, from the same plant (T. corymbosa Roxb. ex Wall).

Criofolinine (1) was initially obtained as a light yellowish oil and subsequently crystallized from absolute ethanol as colorless block crystals, mp >190 °C dec, with $[\alpha]^{25}_{D}$ +87 (CHCl₃, c 0.3). The IR spectrum showed bands due to NH/OH (3393 cm[−]¹) and various carbonyl (1699, 1648 cm[−]¹) functions, while the UV spectrum showed characteristic 2-acylindole absorption maxima at 205, 238, and 316 nm (log ε 4.67, 4.35, and 4.41 respectively).¹⁰ The ESIMS showed an $[M + H]$ ⁺ peak at m/z 399, and HRESIMS measurements $([M + H]^+$ 399.1550) establis[he](#page-3-0)d the molecular formula as $C_{21}H_{22}N_2O_6^{-11}$

The ¹H NMR data (Table 1) showed the presence of four aromatic resonances (δ 7.17–7.61), an indolic [NH](#page-3-0) (δ 8.94), and a methoxy corresponding [to](#page-1-0) a methyl ester group (δ 3.88). The ¹³C NMR data (Table 2) showed a total of 21 carbon resonances, comprising one methyl, four methylene, eight methine, and eight quaternar[y](#page-1-0) carbon atoms. The resonance at

 δ 191.8 was due to a conjugated ketone carbonyl and can be readily assigned to C-3, as it is part of the acyl indole moiety. Two other carbonyl resonances were observed at δ 173.5 and 174.2, which were assigned to ester and lactam carbonyl functionalities, respectively. Assignment of the former resonance to the ester carbonyl was facilitated by the observed three-bond correlation from the ester methyl to the carbonyl resonance at δ 173.5 in the HMBC spectrum. The carbon resonances of the indole unit can be readily assigned based on analogy with other 2-acylindole alkaloids $10a,12$ and these assignments were readily corroborated by NOE and 2D NMR data. A downfield resonance at δ 92.2 [was c](#page-3-0)haracteristic of a quaternary carbon linked to a nitrogen and an oxygen atom,¹³ while another resonance at δ 73.3 was due to an oxymethine.

Received: October 21, 2014 Published: December 2, 2014

Table 1. 1 H NMR Spectroscopic Data (δ) for 1–3 (600 MHz, CDCl₃)

Table 2. ¹³C NMR Spectroscopic Data (δ) for 1–3

The COSY spectrum (Figure 1) showed two partial structures, an NCH_2CH_2 and a CH−CH−CH−CH−CH₂− $CH₂$ fragment, corresponding to a cyclohexane moiety. The

Figure 1. COSY and selected HMBCs of 1 and 3.

assignment of the NCH_2CH_2 fragment to C-5–C-6 was supported by the three-bond correlations from H-6 to C-2, C-8, and from H-5 to C-7, in the HMBC spectrum (Figure 1). The lactam carbonyl was deduced to be linked to N-4, from the observed H-5 to C-21 three-bond correlation. The same applies to the oxygen- and nitrogen-linked C-14 from the observed H-5 to C-14 correlation. The correlation from H-15 to the ketone carbonyl C-3, and from H-16 to C-14, indicated that the carbinol amine C-14 was linked to C-3.

Consideration of the ${}^{1}H$ and ${}^{13}C$ chemical shifts allow the cyclohexane fragment to be rewritten as an $CH(C=O)-CH CH(CO₂Me) – CH(OH) – CH₂–CH₂– corresponding to C-$ 20−C-15−C-16−C-17−C-18−C-19. This six-membered ring E must therefore be linked to the lactam C-21 via C-20 and to the carbinol amine C-14 via C-15, which completes assembly of the $6/5/7/5/6$ pentacyclic ring system of criofolinine (1) .

The relative configurations at the various stereogenic centers were established from the NOE data and the observed vicinal coupling constants. The D/E ring junction stereochemistry was deduced to be *trans* from the observed J_{15-20} value of 12 Hz (H-15 and H-20 trans-diaxial). The reciprocal NOEs observed for H-16/H-18, H-16/H-20, H-18/H-20, and for H-15/H-17, H-15/H-19, H-17/H-19, indicated that these hydrogens are axially oriented, which were consistent with a chair conformation adopted by the E-ring with the OH and $CO₂Me$ substituents equatorially oriented (Figure 2). This was also in agreement with the observed J_{15-16} and J_{16-17} values of 12 and 10 Hz, respectively. The configuration at t[he](#page-2-0) carbinol amine C-14 could not be assigned with certainty based on the spectroscopic data alone but was nonetheless eventually established from X-ray analysis of 1, which also provided confirmation of the structure (Figure 2, relative configuration) of this novel alkaloid deduced from the spectroscopic data.¹⁴

Figure 2. Selected NOEs and X-ray crystal structure of 1.

Criofolinine (1) represents a new monoterpenoid indole alkaloid skeleton, incorporating a pyrroloazepine motif within a pentacyclic ring system.

Vernavosine (2) was isolated as its ethyl ether derivative (3) , which was obtained as a yellow-green fluorescent oil, with $[\alpha]^{25}$ _D −49 (CHCl₃, c 0.31). The UV spectrum showed absorption maxima at 233, 257, and 396 nm, somewhat reminiscent of alkaloids possessing pseudoindoxyl chromophores,^{10b,15} while the IR spectrum showed bands due to OH (3416 cm[−]¹) and various carbonyl (1712 cm[−]¹) functions. The ESIMS sh[owed](#page-3-0) a $[M + H]^+$ peak at m/z 415, and HRESIMS measurements $([M + H]^{\mathrm{+}}_2$ 415.2233) established the molecular formula as $C_{23}H_{30}N_2O_5$.¹⁶

The ¹H NMR spectrum of 3 (Table 1) showed the presence of four aromatic resona[nce](#page-3-0)s associated with the indole moiety (δ 6.81–7.59), a methine linked to [tw](#page-1-0)o nitrogen atoms (δ 4.55), an oxymethine (δ 3.87), a methyl singlet (δ 3.73) due to methyl ester group (δ _C 51.7, 174.7), and an ethoxy side chain $(\delta_{\rm H}$ 1.19, $\delta_{\rm C}$ 14.7; $\delta_{\rm H}$ 3.28, 3.32; $\delta_{\rm C}$ 59.3). The notable absence of the characteristic indolic NH signal indicated substitution at the indolic nitrogen (N-1). The $13C$ NMR data (Table 2) showed a total of 23 carbon resonances, comprising two methyl, seven methylene, nine methine, and five quatern[ary](#page-1-0) carbon atoms. Two carbonyl resonances were observed at δ 200.4 and 174.7, the former was due to a conjugated ketone, while the latter was assigned to the ester carbonyl. The ketone carbonyl was deduced to be at C-7 from the three-bond correlation from H-9 in the HMBC spectrum. In addition, an oxymethine resonance was seen at δ 71.6, while the resonance at δ 89.3 was due to a quaternary carbon linked to a nitrogen, and an oxygen atom.¹³ This carbon corresponded to C-2 to which the ethoxy substituent is linked from the observed threebond correlation fro[m th](#page-3-0)e ethoxy methylene hydrogens (H-24) to this carbon in the HMBC spectrum. The resonance at δ 69.4, which was associated with the $^1\mathrm{H}$ resonance at δ 4.55, provided additional support for the presence of an aminal carbon.

The COSY spectrum showed in addition to the aromatic and ethoxy moieties, two other partial structures, NCH_2CH_2 and NCHCH₂CHCHCHCH₂CH₂CHCH₂ (Figure 1). The former two-carbon fragment corresponded to C-5−C-6 from the threebond correlation from H-5 to C-2 observed [in](#page-1-0) the HMBC spectrum (Figure 1). The nine-carbon fragment corresponded to C-3−C-14−C-15−C-16−C-17−C-18−C-19−C-20−C-21. The aminal carb[on](#page-1-0), C-3 (δ _H 4.55; δ _C 69.4) was linked to both N-1 and N-4, while the assignments of the methyl estersubstituted C-16 and hydroxy-substituted C-17, were consistent with the corresponding carbon resonances observed at δ 57.5 and 71.6, respectively. Similarly for C-21 (δ 59.3), which was linked to N-4. These assignments were in excellent agreement with the full HMBC data (Figure 1). The H-3 to C-13, C-2, and C-5 correlations were consistent with branching of C-3 from N-1 (and N-4), while th[e](#page-1-0) H-21 to C-3 and C-5

correlations were consistent with the connection of C-21 to N-4.

Examination of the vicinal coupling constants $(J_{5\beta\text{-}6\alpha}, J_{3\alpha\text{-}14\beta},$ $J_{14β_15α}$ $J_{15α_16β}$, $J_{16β_17α}$, $J_{17α_18β}$, $J_{18β_19α}$, $J_{19α_20β}$, $J_{20β_21α}$ ~ $11-14$ Hz) and the NOE data (Figure 3), indicated that the C, D, and

Figure 3. Selected NOEs of 3 and X-ray crystal structure of 3a.

E rings adopted the stable chair conformations, with cis-fused C/D and trans-fused D/E rings, and with the C-16 methyl ester and C-17 OH groups oriented equatorially. The C/D cis-ring fusion was also supported from the X-ray diffraction data of the methyl iodide salt of 3 (3a, Figure 3).¹⁷ The ethoxy group was deduced to be β -oriented from the observed H-24/H-14 NOEs and from its presumed origin, whi[ch](#page-3-0) required the alcohol nucleophile to approach the precursor iminium ion from the less hindered β -face (Scheme 1). Vernavosine (2) represents another novel monoterpenoid indole alkaloid skeleton,

characterized by incorporation of a pyridopyrimidine moiety embedded within a pentacyclic ring system.

We propose that both alkaloids originate from a common β yohimbine precursor 4, which was among the alkaloids present in the plant (Scheme 1). Thus, hydrolytic cleavage of the iminium ion 5 derived from oxidation of the $β$ -yohimbine precursor 4 gave the k[eto](#page-2-0) amine 6. Reduction of the ketone function, followed in succession by dehydration and oxidation, yielded the epoxide 7. Epoxide ring opening via transannular attack by the secondary amine nitrogen forged the pyrroloazepine ring system of the alcohol 8, which on selective oxidation of the benzylic alcohol moiety gave the conjugated ketone 9. Nucleophilic attack by water on the iminium ion derived from 9 installed the tertiary alcohol functionality at C-14, and a final oxidation provided criofolinine (1). Alternatively, oxidation of the same $β$ -yohimbine precursor 4 gave the pseudoindoxyl alkaloid 10. A further oxidation provided the N(4)-oxide derivative 11, which on a lone-pair assisted Groblike fragmentation (Polonovski-like) yielded the imine-iminium ion intermediate 12. Ring closure via attack of the imine nitrogen (N-1) on the iminium ion forged the new pentacyclic ring system of vernavosine in the form of its iminium ion 13, which on reaction with water yields the carbinol amine 2. In the presence of the stronger ethanol nucleophile,¹⁸ the carbinol amine 2 will in all probability be readily converted to its ethanolysis product 3, which was the final form of the alkaloid isolated. Hydrolysis of 3 (in two-phase medium with phasetransfer catalysis) gave the putative precursor alkaloid, the carbinol amine 2, while re-exposure of 2 to EtOH in the presence of a trace of acid gave 3, providing additional confirmation for the origin of the ethyl ether derivative 3 from the original intact alkaloid 2.¹⁹

Both compounds 1 and 3 showed no appreciable cytotoxicity against drug-sensitive as well as drug-resistant KB cells, HCT-116, PC-3, and A-549 cells (IC₅₀ > 25 μ g mL⁻¹ or 60 μ M). Compound 3 however, showed a moderate concentration dependent relaxation effect on phenylephrine-induced contraction in isolated rat aortic rings with $EC_{50} = 2.48 \mu M$ and E_{max} = 39.4 \pm 4.4% (cf. isoprenaline, EC₅₀ = 0.07 μ M and E_{max} $= 79.7 \pm 4.2\%$).²⁰

■ ASSOCIATED CONTENT

S Supporting Information

Experimental procedures, NMR spectra, HRESIMS (1−3), and X-ray crystallographic data (CIF) of 1 and 3a. This material is available free of charge via the Internet at http://pubs.acs.org.

■ AUTHOR INFORMATION

Corresponding Author

*E-mail: tskam@um.edu.my.

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

We thank the University of Malaya and MOHE Malaysia (HIR-005) for financial support and Dr. K. S. Sim, Institute of Biological Sciences, University of Malaya, for the cytotoxicity assays.

■ REFERENCES

(1) (a) Leeuwenberg, A. J. M. Tabernaemontana: The Old World Species; Royal Botanic Gardens, Kew: UK, 1991.

(2) (a) Van Beek, T. A.; Verpoorte, R.; Svendsen, A. B.; Leeuwenberg, A. J. M.; Bisset, N. G. J. Ethnopharmacol. 1984, 10, 1−156. (b) Danieli, B.; Palmisano, G. In The Alkaloids; Brossi, A., Ed.; Academic: Orlando, 1986; Vol. 27, pp 1−130. (c) Van Beek, T. A.; Van Gessel, M. A. J. T. In Alkaloids: Chemical and Biological Perspectives; Pelletier, S. W., Ed.; Wiley & Sons: New York, 1988; Vol. 6, pp 75−226.

(3) (a) Kam, T. S. In Alkaloids: Chemical and Biological Perspectives; Pelletier, S. W., Ed.; Pergamon: Amsterdam, 1999; Vol. 14, pp 285− 435. (b) Kam, T. S.; Choo, Y. M. In The Alkaloids; Cordell, G. A., Ed.; Academic Press: Amsterdam, 2006; Vol. 63, pp 181−337.

(4) (a) Kam, T. S.; Sim, K. M.; Lim, T. M. Tetrahedron Lett. 2000, 41, 2733−2736. (b) Kam, T. S.; Sim, K. M.; Lim, T. M. Tetrahedron Lett. 2001, 42, 4721−4723. (c) Lim, K. H.; Etoh, T.; Hayashi, M.; Komiyama, K.; Kam, T. S. Tetrahedron Lett. 2009, 50, 752−754. (d) Lim, K. H.; Kam, T. S. Tetrahedron Lett. 2009, 50, 3756−3759. (5) (a) Lim, K. H.; Hiraku, O.; Komiyama, K.; Kam, T. S. J. Nat. Prod. 2008, 71, 1591−1594. (b) J. Raja, V.; Lim, K. H.; Leong, C. O.; Kam, T. S.; Bradshaw, T. D. Invest. New Drugs 2014, 32, 838−850. (c) Frei, R.; Staedler, D.; Raja, A.; Franke, R.; Sasse, F.; Gerber-Lemaire, S.; Waser, J. Angew. Chem., Int. Ed. 2013, 52, 13373−13376. (6) (a) Kam, T. S.; Sim, K. M.; Koyano, T.; Toyoshima, M.; Hayashi, M.; Komiyama, K. Bioorg. Med. Chem. Lett. 1998, 8, 1693−1696. (b) Kam, T. S.; Sim, K. M.; Pang, H. S. J. Nat. Prod. 2003, 66, 11−16. (7) (a) Kam, T. S.; Loh, K. Y.; Lim, L. H.; Loong, W. L.; Chuah, C. H.; Chen, W. Tetrahedron Lett. 1992, 33, 969−972. (b) Kam, T. S.; Loh, K. Y.; Chen, W. J. Nat. Prod. 1993, 56, 1865−1871. (c) Kam, T. S.; Pang, H. S.; Lim, T. M. Org. Biomol. Chem. 2003, 1, 1292−1297. (8) (a) Umezawa, K.; Ohse, T.; Yamamoto, T.; Koyano, T.; Takahashi, Y. Anticancer Res. 1994, 14, 2413−2418. (b) Umezawa, K.; Hiroki, A.; Kawakami, M.; Naka, H.; Takei, I.; Ogata, T.; Kojimi, I.; Koyano, T.; Kowithayakorn, T.; Pang, H. S.; Kam, T. S. Biomed. Pharmacother. 2003, 57, 341−350.

(9) Nge, C. E.; Gan, C. Y.; Low, Y. Y.; Thomas, N. F.; Kam, T. S. Org. Lett. 2013, 15, 4774−4777.

(10) (a) Kam, T. S.; Pang, H. Y.; Choo, Y. M.; Komiyama, K. Chem. Biodiversity 2004, 1, 646−656. (b) Sangster, A. W.; Stuart, K. L. Chem. Rev. 1965, 65, 69−130.

(11) HRESIMS found m/z 399.1550 [M + H]⁺ (calcd for $C_{21}H_{22}N_2O_6 + H$, 399.1551).

(12) Verpoorte, R.; Van Beek, T. A.; Riegman, R. L. M.; Hylands, P. J.; Bisset, N. G. Org. Magn. Reson. 1984, 22, 328−335.

(13) (a) Gan, C. Y.; Low, Y. Y.; Thomas, N. F.; Kam, T. S. J. Nat. Prod. 2013, 76, 957−964. (b) Kam, T. S.; Subramaniam, G.; Lim, K. H.; Choo, Y. M. Tetrahedron Lett. 2004, 45, 5995−5998.

(14) The crystals of 1 are orthorhombic, belonging to space group $P2_12_12_1$, with $a = 10.7249(5)$ Å, $b = 12.1181(6)$ Å, $c = 29.2040(13)$ Å, $V = 3795.5(3)$ Å³, T = 150 K, $D_{\text{calcd}} = 1.394$ mg/mm³, and Z = 4. The final R₁ value is 0.0492 (wR₂ = 0.1440) for 7804 reflections [I > $2\sigma(I)$]. CCDC No. 1029243.

(15) Stahl, R.; Borschberg, H. J. Helv. Chim. Acta 1994, 77, 1331− 1345.

(16) HRESIMS found m/z 415.2233 $[M + H]^+$ (calcd for $C_{23}H_{30}N_2O_5 + H$, 415.2227).

(17) The crystals of 3a are monoclinic, belonging to space group P_1 , with $a = 10.4239(2)$ Å, $b = 8.4186(2)$ Å, $c = 14.8476(3)$ Å, $V =$ 1273.54(5) Å³, T = 150 K, $D_{\text{caled}} = 1.451 \text{ mg/mm}^3$, and Z = 2. The final R_1 value is 0.0382 (w R_2 = 0.1011) for 5866 reflections [I > $2\sigma(I)$]. Flack parameter $[x = 0.018(15)]$, Hooft parameter $[y =$ 0.008(17)]. CCDC No. 1029244.

(18) EtOH was used during extraction of alkaloids.

(19) Compound 2: $[\alpha]_{D}^{25}$ –62 (CHCl₃, c 0.06); HRESIMS found m/z 387.1914 $[M + H]^+$ (calcd for $C_{21}H_{26}N_2O_5 + H$, 387.1914); UV (EtOH), λ_{max} (log ε) 234 (3.48), 2.57 (2.91), 397 (2.59) nm ; ¹H and ¹³C NMR, see Tables 1 and 2.

(20) (a) Tsai, T. H.; Wang, G. J.; Lin, L. C. J. Nat. Prod. 2008, 71, 289−291. (b) Matsuo, H.; Okamoto, R.; Zaima, K.; Hirasawa, Y.; Ismail, I. S.; Lajis, N. [H.;](#page-1-0) Mo[rit](#page-1-0)a, H. Bioorg. Med. Chem. Lett. 2011, 19, 4075−4079.